

From Page No. 37

Checked mammalian fusion construct transformation plates

Started 40 x 5ml CB cant 50 (+ master) MP's

Inc all 37°C O/N.

Want to prepare HPTKb full length in pSV15.1

need to prepare EcoRI/BamHI vector

RD <sup>+</sup> mixed	10µl	vector DNA
	10µl	10x H
	75µl	H <sub>2</sub> O
	5µl	EcoRI
	100	

Inc 37°C O/N.

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Witnessed &amp; Understood by me,

Date

Invented by

Da

H. H. H. Bacon

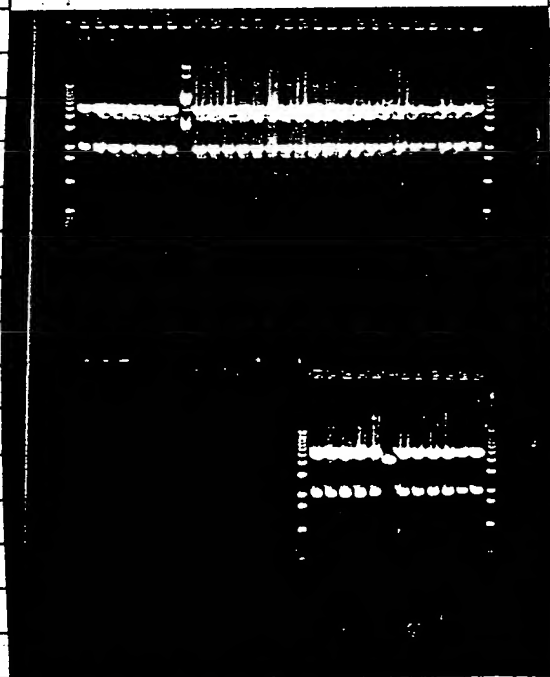
TITLE \_\_\_\_\_

From Page No. 38

Ran 40 O/N MP cultures in Autogen Masters to  
Recovered each in ~200  $\mu$ l TE

RD's

Per rxn

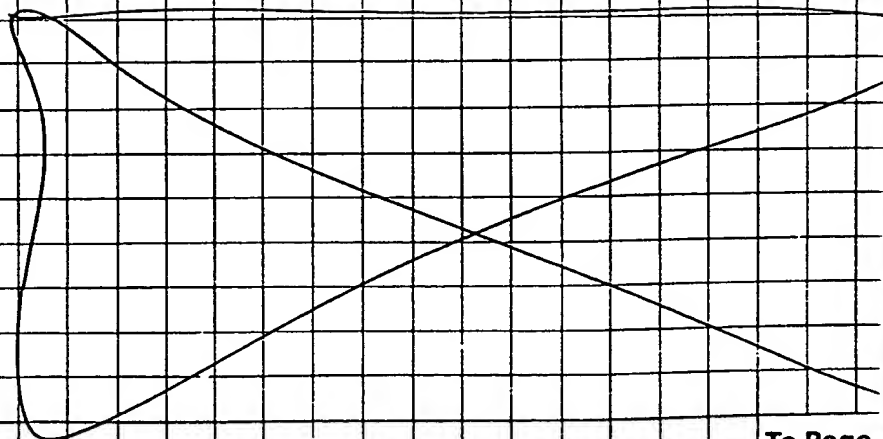
4  $\mu$ l MP DNA2  $\mu$ l 10x B0.5  $\mu$ l EcoRI0.5  $\mu$ l HindIII1  $\mu$ l RNase A12  $\mu$ l H<sub>2</sub>O20Inc 37°C ~1.5 hrs  $\rightarrow$  added 4  $\mu$ l dye to eachRan 15  $\mu$ l each on 0.7% agarose (1x TBE)

only 2 negatives

Started 2x 500ml 2YT cult-50

Maxi's on #1 &amp; #2

Inc 37°C O/N



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From Page No. 39

Want to subclone full length HPTK6 to pSV15-ID-11  
for stable transfection.

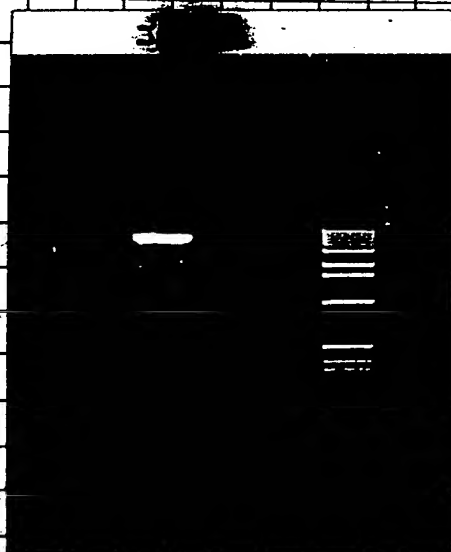
(Have R1/BamHI insert already prepared)

Vector digest:

mixed 4  $\mu$ l MP DNA (pSV15) (~1.5  $\mu$ g)  
2  $\mu$ l 10x B  
1  $\mu$ l BamHI  
2  $\mu$ l EcoRI  
12  $\mu$ l H<sub>2</sub>O  
20

Inc 37°C 1.5 hrs  $\rightarrow$  added 4  $\mu$ l dye

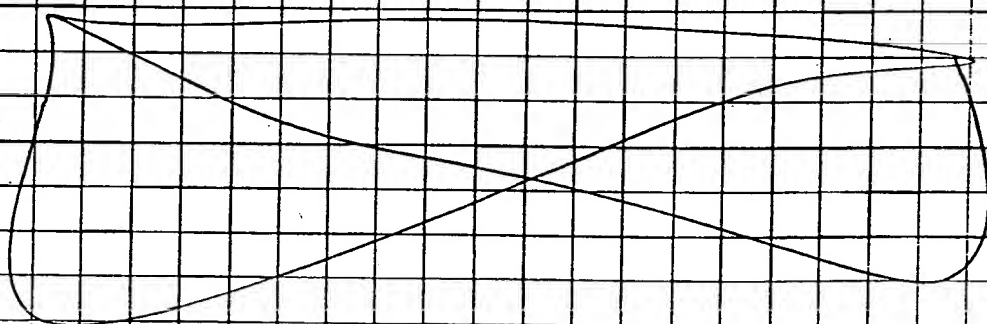
Ran entire rxn on 1% agarose (1x TBE)



cut out indicated band  
Did Magic PCR prep  
R<sub>5</sub>'d in 150  $\mu$ l TE

Ligated to HPTK6 (Hi & Lo conc)  
(plus vect. cont).

12.5°C o/n.



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NHA Bacon

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Project No. 171

Book No. 17

Exhibit C, pg. 4 of 6

From Page No. 40

Transferred O/W Ciga to  $-20^{\circ}\text{C}$ .

Stored max prep cultures  $4^{\circ}\text{C}$

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1. 11 / 2012

Date

[Redacted]

om Page No. 41

Transformed pSV15/HPTK6 (full length) into  
competent E. coli

Plated each onto 5 x 100mm LB canb 50  
(control onto 1 plate)

Inc all (11) o/n 37°C.

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From Page No. 4

Did magic MP's on 8 o/n pSV15 MP's (Master stored 4°C)

RD's      2µl MP DNA  
            2µl 10x B  
            0.5µl BamH.I  
            0.5µl EcoR.I  
            15µl H<sub>2</sub>O  
            20

Inc 37°C ~ 1.5 hrs → added 4µl dye to each

Ran 15µl each on 0.7% agarose (1x TBE)



5 of 8 positive.

Started 1x 500ml Lb. cult<sup>50</sup> o/n min.

Inc 37°C o/n.

Received M. Mark's Northern Blot  
(+ clontech mouse MTN)Prehybridized according to  
Clontech's protocol  
42°C ~ 6 hrs.Did magic clean-up on o/n SacI  
RD of pGEM-37 HPTK6

Did SalI RD

Ran on 1% LMP agarose (1x TBE)

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Invented by

NIA Baron

Date